

Muonium Chemistry at Diiron Subsite Analogues of [FeFe]-Hydrogenase

Joseph A. Wright,* Jamie N. T. Peck, Stephen P. Cottrell, Aušra Jablonskytė, Vasily S. Oganessian, Christopher J. Pickett,* and Upali A. Jayasooriya*

Abstract: The chemistry of metal hydrides is implicated in a range of catalytic processes at metal centers. Gaining insight into the formation of such sites by protonation and/or electroreduction is therefore of significant value in fully exploiting the potential of such systems. Here, we show that the muonium radical (Mu^\bullet), used as a low isotopic mass analogue of hydrogen, can be exploited to probe the early stages of hydride formation at metal centers. Mu^\bullet undergoes the same chemical reactions as H^\bullet and can be directly observed due to its short lifetime (in the microseconds) and unique breakdown signature. By implanting Mu^\bullet into three models of the [FeFe]-hydrogenase active site we have been able to detect key muoniated intermediates of direct relevance to the hydride chemistry of these systems.

Developing new approaches to gaining insight into catalytic systems is of central importance. We are now exploring the exciting possibility of using muonium radicals as surrogates for H^\bullet in the study of catalytic and electrocatalytic reactivity at metal centers. The system we have chosen is the active site of the [FeFe]-hydrogenase.

The [FeFe]- and [NiFe]-hydrogenases catalyze the reversible reduction of protons to dihydrogen at high turnover frequencies and at low overpotentials. On a per catalytic site basis, the turnover frequencies of certain of these enzymes immobilized on electrodes can rival the best conventional electrocatalyst, carbon-supported platinum.^[1–3] However, the high molecular mass and large geometric footprint of the native enzymes result in rather low current densities, at best ca. 3 mA cm^{-2} at 20°C . Given that hydrogen fuel or producer cells typically operate at current densities some two or three orders of magnitude greater, it is arguable whether hydrogenase-based electrodes are likely to be useful materials in practical devices. Nevertheless, it is well established that the relatively small metallosulfur centers within these enzymes are responsible for their high catalytic activity and this has

prompted extensive research on synthetic analogues of these active sites.^[1,3,4] This research is driven both by the need to provide mechanistic insights into the biological chemistry and the possibility of creating new materials for electrocatalysis based on abundant 3d metals.

Mechanistically, the formation of more or less transient hydride intermediates is central to both hydrogen evolution or uptake by the [FeFe]- and [NiFe]-hydrogenases and electrocatalysis by synthetic analogues of their active sites.^[5] We now describe the first application of muon spin spectroscopy to probe hydride chemistry at metallosulfur sites related to that within a hydrogenase, specifically the diiron subsite of [FeFe]-hydrogenase.

When an energetic muon (μ^+) beam passes through a solid sample some of the sub-atomic μ^+ ions capture an electron to form muonium radicals (Mu^\bullet);^[6] these are sufficiently slowed to react with the bulk target species, in our case the diiron subsite analogue. Muonium can be considered chemically as acting as a light isotope of hydrogen atom, having about 1/9 of its mass; Mu^\bullet thus provides a surrogate for hydrogen radicals. The implantation of a muonium radical on a diiron-subsite analogue can be viewed as the equivalent of concerted addition of a proton and an electron; it takes place on the nano- to microsecond timescale giving rise to paramagnetic species.^[6] Analogous to the proton, the muon has a nuclear spin of one-half; at the end of its life (lifetime $2.197 \mu\text{s}$) it decays to give a positron which is emitted preferentially along the spin direction at the moment of decay. Because it is possible to produce almost 100% spin-polarized muon beams, detection of the direction of emission of the decay positrons allows the study of the evolution of the muon(ium) spin within the implanted sample. This gives information on the system under scrutiny via the magnetic fields local to the muon site, μSR (muon spin rotation/relaxation/resonance) spectroscopy.^[7]

To date, there have been only a very small number of studies on application of μSR to organometallic systems.^[8] These have been focused on silylenes,^[9] or cyclopentadienyl or arene (half)sandwich systems;^[10–13] in these compounds protonation and proton-coupled electron transfer are of limited relevance to the key chemical reactions of the metal complexes. In contrast, the spectroscopic data we have so obtained can be modelled by ab initio DFT calculations and we show that this provides compelling evidence for the muono-formyl $\text{Fe-C}(\text{Mu})=\text{O}$ and bridging Fe-Mu-Fe or terminal Fe-Mu muonide transients, light isotopes of paramagnetic species that are implicit in hydrogen evolution and uptake.^[1–3,14]

[*] Dr. J. A. Wright, Dr. J. N. T. Peck, Dr. A. Jablonskytė, Dr. V. S. Oganessian, Prof. C. J. Pickett, Dr. U. A. Jayasooriya
Energy Materials Laboratory, School of Chemistry,
University of East Anglia, Norwich, NR4 7TJ (UK)
E-mail: joseph.wright@uea.ac.uk
c.pickett@uea.ac.uk
u.jayasooriya@uea.ac.uk

Dr. J. N. T. Peck, Dr. S. P. Cottrell
Rutherford Appleton Laboratory
Harwell Oxford, Didcot OX11 0QX (UK)

Supporting information and the ORCID identification number(s) for the author(s) of this article can be found under <http://dx.doi.org/10.1002/anie.201607109>.

As far as we are aware, there are no studies using μ SR spectroscopy to probe chemical processes relevant to metal hydrides in catalysis or electrocatalysis. In this work, we have examined three [FeFe]-hydrogenase subsite models, each of which illustrate a different aspect of hydride chemistry (Figure 1). Thus, complex **1** is known to engage in electro-

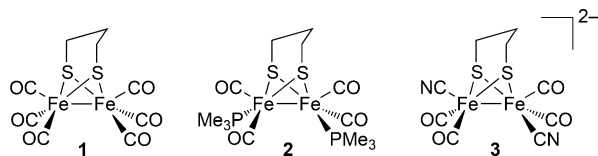


Figure 1. Complexes 1–3.

catalysis, in which an electron and a proton are added successively;^[15] complex **2** protonates at the metal–metal bond^[16] enabling subsequent electronation to yield a mixed-valence Fe(1.5)Fe(1.5) hydride (cf. H-atom addition);^[17] whilst complex **3** possesses the bis-cyanide coordination found in the enzyme and has been shown to reconstitute an apoenzyme.^[18–20]

In the solid state (an anisotropic environment), coupling between the radical (electron) and muon spins gives rise to the Δ_1 transition.^[6] This may be probed using “avoided level crossing muon spin resonance” (ALC- μ SR).^[6] ALC is a longitudinal field μ SR technique which detects the reduction in polarization at “level crossings” in the Breit–Rabi diagram. For Δ_1 resonances, where only the muon spin changes sign, the resonance field is related to the hyperfine interaction by equation (1).

$$B_{\text{res}}^{\Delta_1} = \frac{1}{2} \left[\frac{A_{\mu}}{\gamma_{\mu}} - \frac{A_{\text{c}}}{\gamma_{\text{c}}} \right] \quad (1)$$

ALC- μ SR spectra of **1–3** at 300 K over the field range 2 kG to 18 kG are shown in Figure 2.^[21] The general features of all three spectra are remarkably similar: each room temperature spectrum exhibits a broad signal with a maximum in the range 8 kG to 10 kG. The signals all show significant temperature dependence, with essentially complete loss of the signal at the lowest temperature (10 K) for **1** and **3**, whilst **2** exhibits only a small residual signal.

Given the very broad nature of the ALC- μ SR signals, accurate modelling of the background is essential. Scanning using a cell containing a mass of aluminum foil equivalent to the sample showed a smooth curve which could be fitted using a fourth-order polynomial (Figure S1). Whilst the line shape for the solid-state anisotropic signals here is expected to be complex, it is possible to approximate the spectra using Gaussian curves following subtraction of the cell background (Figure 3, Figures S5, S6). This process confirms the presence of a common major signal at around 8.5 kG in all three samples. A second maximum at ca. 4.5 kG can clearly be seen at 300 K for **2** and at lower temperatures for **1** (Figure 2, Figures S5, S6). Complex **3** does not show this signal.

The specific nature of the muonium species which give rise to the ca. 8.5 kG and ca. 4.5 kG signals has been probed

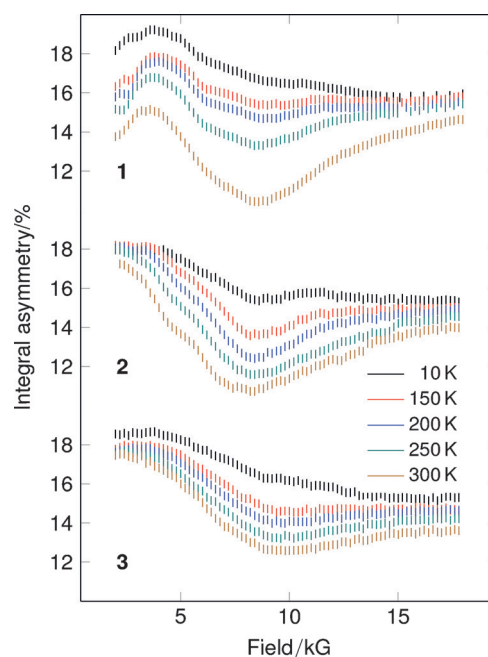


Figure 2. Raw time-integral ALC- μ SR spectra for **1–3**. Data points are shown as sticks representing the estimated uncertainty in each point. The vertical scale is the same for each spectrum.

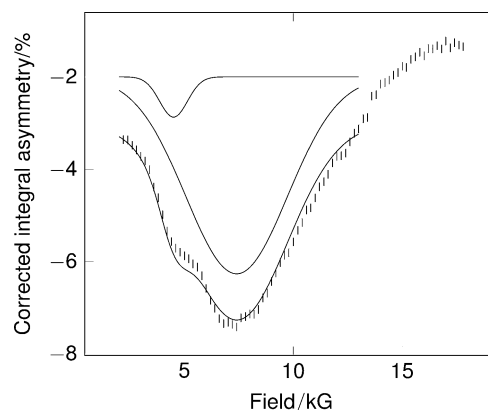


Figure 3. Background-subtracted time-integral ALC- μ SR spectra for **2** at 300 K showing Gaussian line shape approximation for the range 2 kG to 13 kG.

by computational simulation of plausible structures, from which Δ_0 values have been calculated.^[22] For the known solid-state structure of **2** which has a basal–basal deployment of phosphine ligands,^[16] the high-field signal at 8.5 kG is consistent with the Mu being bound to iron either in a bridging or terminal position (Figure 4). For **3**, only the bridging muonide fits with the experimental value. The low-field signal at 4.5 kG can be accounted for by formation of a formyl-like radical (Figure 5, left). We note that structures in which the rotation of a tripodal ligand group has occurred can also accommodate the observed signals (Figure 5, right), but such a rotation is perhaps unlikely in the solid state given the timeframe of the ALC- μ SR experiment.

It is important to note that **2-Mu_{bridge}** is structurally analogous to the mixed-valence bridging hydride detected by

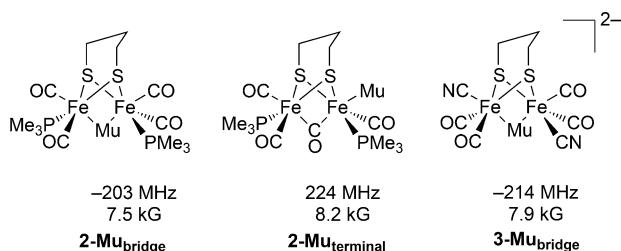


Figure 4. Potential sites of muon addition, and calculated hyperfine coupling and resonance field values.

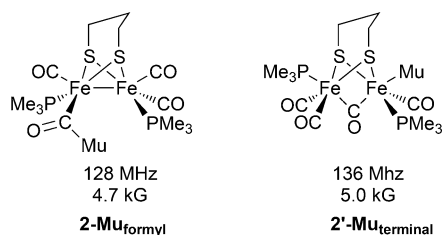


Figure 5. Example sites with resonance fields of around 5.0 kG.

electron paramagnetic resonance on one-electron reduction of the closed-shell cationic hydride, $[\text{HFe}_2(\text{pdt})(\text{CO})_4(\text{PMe}_3)_2]^+$,^[17] and that **1-Mu_{formyl}** can be regarded as an isotopomer of the formyl species observed upon reduction of **1** in presence of acid.^[6,15]

Figure 2 shows that the intensities of the signals are temperature-dependent. At low temperature (10 K) the response approaches that of the cell background. Figure 6 shows the Arrhenius plot for the major resonance in each of the three samples where the 10 K data is subtracted with the

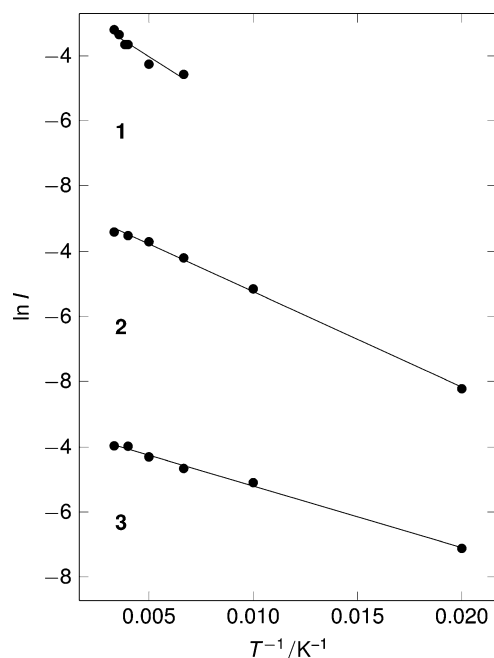


Figure 6. Arrhenius plots for signal intensity following muonation, subtracting 10 K spectra and correcting for temperature-dependent drift. (Resonance positions: **1** 8.4 kG, **2** 8.2 kG, **3** 10.0 kG).

spectra pinned to zero at 14 kG (**1**)/18 kG (**2** and **3**). For the full range of data **2** and **3** there is an excellent linear correlation of $\ln I$ with $1/T$.^[23] The more limited data for **1** shows a similar trend. The estimated activation energies obtained for the addition at the bridging position are **1** 3.40(2) kJ mol⁻¹, **2** 2.45(2) kJ mol⁻¹ and **3** 1.58(3) kJ mol⁻¹, consistent with the high reactivity of muonium. The order obtained is notable, with activation energies falling as the systems become more electron rich: **1** is the least electron rich, **3** the most.

The second resonance which we have assigned to **Mu_{formyl}** in the data for the PMe_3 complex (**2**) remains resolved across the temperature range 100 K to 300 K. From the raw $\ln I$ and $1/T$ data we estimate an activation energy of ca. 9 kJ mol⁻¹ from the Arrhenius relationship (correlation coefficient 0.965). In the lower temperature scans, the intensity of the resonance is lost in the background.

As a further verification that radical states are formed, repolarization experiments were also carried out (Figure 7, Figure S14). We clearly observed the recovery of polarization across the temperature range. Whilst such experiments confirm radical generation, extraction of hyperfine coupling

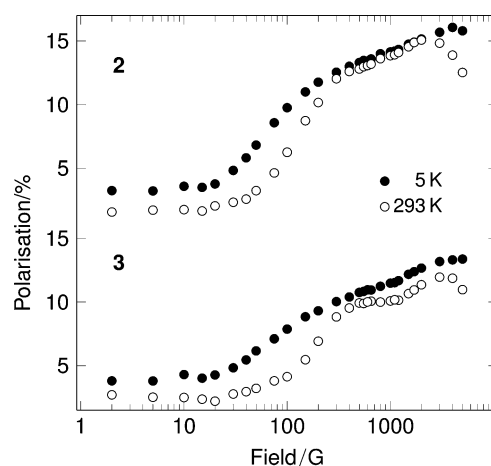


Figure 7. Repolarization spectra for **2** and **3** at ambient and base temperatures. Estimated uncertainty values were smaller than the markers used for all points. The vertical scale is the same for both spectra.

constants is known to be difficult.^[6] It is notable however that there is recovery at the lowest temperature (5 K), where the ALC- μSR spectra are essentially featureless. Thus, radical states are still being formed at low temperature but likely broadened beyond detection in the ALC- μSR . This may be due to the existence of multiple sites of addition, consistent with the DFT calculations, and/or may be due to electron-spin relaxation.

In summary, we have provided evidence that muonium interacts in the solid state with the diiron subsite analogues **1**–**3** to give 35-electron bridging muonides $(\mu\text{-Mu})\text{Fe}_2(\text{pdt})(\text{CO})_4(\text{L})_2$. The formation of these species is temperature-dependent with activation energies less than 4 kJ mol⁻¹. In the case $\text{L} = \text{PMe}_3$ (**2**) we observed a second resonance, assigned

to the formation of a muono-formyl species, -COMu. The generation of these muoniated species have direct parallels in the protonation/electronation of substrates^[17] and represent the first example of the application of μ SR to electrocatalytic systems.

The work described here is likely to presage a wider application of muonium chemistry. The role of metal-hydride interactions in diverse inorganic, organometallic, and biological chemistry is extensive, ranging from β -elimination through water-gas shift chemistry, the chemistry of the hydrogenases and nitrogenases, to electrocatalytic systems for hydrogen fuel/producer cells. Muonium chemistry coupled with the fast timescale of μ SR spectroscopy offers the prospect of unravelling mechanistic detail of such systems, for example, the possibility of detecting transient dihydrogen/dihydride analogues.

Acknowledgments

This research was supported by the EPSRC (grant EP/M011879/1) and the University of East Anglia. Experiments at the ISIS Pulsed Neutron and Muon Source were supported by a beamtime allocation from the Science and Technology Facilities Council.

Keywords: enzyme mimics · [FeFe]-hydrogenase · hydrogen · muonium

How to cite: *Angew. Chem. Int. Ed.* **2016**, 55, 14580–14583
Angew. Chem. **2016**, 128, 14800–14803

- [1] C. Tard, C. J. Pickett, *Chem. Rev.* **2009**, 109, 2245–2274.
- [2] W. Lubitz, H. Ogata, O. Rüdiger, E. Reijerse, *Chem. Rev.* **2014**, 114, 4081–4148.
- [3] T. R. Simmons, G. Berggren, M. Bacchi, M. Fontecave, V. Artero, *Coord. Chem. Rev.* **2014**, 270–271, 127–150.
- [4] F. Gloaguen, T. B. Rauchfuss, *Chem. Soc. Rev.* **2009**, 38, 100–108.
- [5] N. T. Hunt, J. A. Wright, C. J. Pickett, *Inorg. Chem.* **2016**, 55, 399–410.
- [6] I. McKenzie, *Annu. Rep. Prog. Chem. Sect. C* **2013**, 109, 65–112.
- [7] U. A. Jayasooriya, R. Grinter in *Encyclopaedia of Applied Spectroscopy* (Ed.: D. L. Andrews), Wiley-VCH, Weinheim, **2009**, pp. 153–182.
- [8] For studies of organic radicals see: C. J. Rhodes, *J. Chem. Soc. Perkin Trans. 2* **2002**, 1379–1396. For insight into the kinetics of muon addition to small molecules, see for example: D. J. Arseneau, D. M. Garner, I. D. Reid, D. G. Fleming, *J. Phys. Chem. A* **2015**, 119, 7247–7256; J.-C. Brodovitch, B. M. McCollum, P. W. Percival, *Physica B: Condens. Matter* **2009**, 404, 950–952.
- [9] R. West, P. W. Percival, *Dalton Trans.* **2010**, 39, 9209–9216.
- [10] U. A. Jayasooriya, J. A. Stride, G. M. Aston, G. A. Hopkins, S. F. Cox, S. P. Cottrell, C. A. Scott, *Hyperfine Interact.* **1997**, 106, 27–32.
- [11] U. A. Jayasooriya in *Fluxional Organometallic and Coordination Compounds* (Eds.: M. Gielen, R. Willem, B. Wrackmeyer), Wiley, Chichester, **2004**, pp. 243–265.
- [12] U. A. Jayasooriya, R. Grinter, P. L. Hubbard, G. M. Aston, J. A. Stride, G. A. Hopkins, L. Camus, I. D. Reid, S. P. Cottrell, S. F. J. Cox, *Chem. Eur. J.* **2007**, 13, 2266–2276.
- [13] I. McKenzie, *Phys. Chem. Chem. Phys.* **2014**, 16, 10600–10606.
- [14] J. A. Wright, C. J. Pickett in *Bioinspired Catalysis* (Eds.: W. Weigand, P. Schollhammer), Wiley-VCH, Weinheim, **2014**, pp. 161–198.
- [15] S. Borg, T. Behrsing, S. P. Best, M. Razavet, X. Liu, C. J. Pickett, *J. Am. Chem. Soc.* **2004**, 126, 16988–16999.
- [16] X. Zhao, I. P. Georgakaki, M. L. Miller, J. C. Yarbrough, M. Y. Darensbourg, *J. Am. Chem. Soc.* **2001**, 123, 9710–9711.
- [17] A. Jablonskytė, J. A. Wright, S. A. Fairhurst, J. N. T. Peck, S. K. Ibrahim, V. S. Oganessian, C. J. Pickett, *J. Am. Chem. Soc.* **2011**, 133, 18606–18609.
- [18] F. Gloaguen, J. D. Lawrence, T. B. Rauchfuss, *J. Am. Chem. Soc.* **2001**, 123, 9476–9477.
- [19] J. A. Wright, L. R. Webster, A. Jablonskytė, P. M. Woi, S. K. Ibrahim, C. J. Pickett, *Faraday Discuss.* **2011**, 148, 359–371.
- [20] G. Berggren, A. Adamska, C. Lambert, T. R. Simmons, J. Esselborn, M. Atta, S. Gambarelli, J.-M. Mouesca, E. Reijerse, W. Lubitz, T. Happe, V. Artero, M. Fontecave, *Nature* **2013**, 499, 66–69.
- [21] Given the very broad nature of the signals seen in the 2 kG to 18 kG region, we explored the possibility of additional resonances in the region extending to 30 kG. Whilst this region is not featureless, background-subtraction of the spectra obtained suggest that there are unlikely to be significant signals in this region (Figures S2 to S4).
- [22] Calculated values for Δ_0 resonances (coupling between the muon spin and the ^{31}P centers) were also considered but yield hyperfine values which would give signals well outside the observed positions.
- [23] The intensity of the resonance (the absolute value of the amplitude of the signal) is given the symbol I here.

Received: July 22, 2016

Published online: October 14, 2016